**REVIEW ARTICLE** 

# ANTIVIRAL COMPOUNDS IN THE PIPELINE TO TACKLE H1N1 INFLUENZA INFECTION

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## **SUMMARY**

The recent pandemic of H1N1 has demonstrated the potential vulnerability of the human population to novel influenza viruses. While there is recent increased interest and effort in developing effective anti-influenza agents, few new products have entered clinical studies. This review will highlight the limited armamentarium of licensed influenza agents, and discuss novel compounds and strategies that have entered clinical studies and may therefore be imminently available to the treating clinician.

#### INTRODUCTION

In March 2009, a novel strain of influenza (A/H1N1 2009) emerged, causing an increase in reports of influenza-like illness in North America. The initial reported cases occurred in Mexico City, which was seen as the epicenter of the influenza pandemic, the world's first in 40 years. Initial reports suggested a high mortality rate (6-7%), although subsequent analysis suggested that these early reports likely inaccurately attributed deaths to novel H1N1, while substantially underestimating the total number of individuals infected with

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mild cases of the disease. As of May 7, 2010, more than 214 countries and overseas territories or communities worldwide have reported laboratory-confirmed cases of pandemic influenza A/H1N1 2009, including at least 18,001 deaths (1). In contrast to seasonal influenza, influenza A/H1N1 2009 affects a younger, healthier population.

Viral glycoproteins (hemagglutinin [HA] and neuraminidase [NA]) are the main targets of the immune response to influenza. Vaccines elicit an immune response to these glycoproteins, but as these vary continuously as a result of antigenic drift and shift, and given the time to produce an antigenically appropriate immunogenic product, antiviral drugs are the principal countermeasure to emerging influenza viruses (2, 3). The currently licensed anti-influenza medications consist of four drugs: two oral adamantanes, amantadine and rimantadine, and two NA inhibitors (NAIs), the oral drug oseltamivir (Tamiflu®) and the inhaled medication zanamivir (Relenza®). Ribavirin, while not licensed for influenza, is available in many countries and does have anti-influenza activity.

#### RESISTANCE

The resistance of influenza A viruses to amantadine or rimantadine can occur spontaneously or emerge rapidly during treatment. Most (> 95%) seasonal H3N2 isolates are resistant to these compounds (4), although they still maintain sensitivity to oseltamivir and zanamivir. Most (> 98%) seasonal H1N1 isolates prior to 2009 were resistant to oseltamivir (5), although they remained sensitive to amantadine, rimantadine and zanamivir. The influenza A/H1N1 2009 isolates are resistant to amantadine and rimantadine (6), and some isolates have additional resistance to oseltamivir and peramivir (7).

#### **DEVELOPMENT OF NOVEL ANTI-INFLUENZA AGENTS**

The limited armamentarium of effective anti-influenza agents and increasing resistance highlight the potential vulnerability of the human population to novel influenza viruses. This has not been overlooked in recent years, as there have been over 2,800 patent publications for aspects of therapeutic anti-influenza agents (8). The development of anti-influenza agents has occurred in the following areas:

- Adamantane-like compounds (muscarinic acetylcholine M<sub>2</sub> receptor antagonists)
- · Antisense, siRNA and miRNA
- Apoptosis inhibitors
- HA antagonists
- Interferons and interleukins as immunomodulators
- Immune hyperactivation antagonists
- NA inhibitors (NAIs)
- NS1 antagonists
- Nucleotide analogues (polymerase inhibitors)
- Monoclonal antibodies
- Polyclonal neutralizing antibodies (serum and immune globulin [IVIG])
- Small-molecule antagonists of cap-dependent viral endonuclease
- RAF/MEK/ERK inhibitors and kinase modulators

Most compounds and strategies involving these approaches are still in preclinical studies, and only a few have entered clinical studies. Figure 1 shows the mechanism of action of the major compounds/ classes in clinical studies. This review will focus on compounds and strategies that have entered clinical evaluation and may therefore be imminently available to the treating clinician. Furthermore, as history suggests that influenza pandemics recur several times a century, strategies that can be applied to both H1N1 and other emerging influenza strains will be reviewed.

## **NEW STRATEGIES WITH CURRENTLY AVAILABLE ANTIVIRALS**

The antiviral strategies that are most easily implemented for the novel H1N1 influenza virus are those that utilize existing licensed antivirals.

## **Dosing strategies**

The trials supporting the use of oseltamivir in the treatment of adults with human influenza who presented within 36 h of developing fever, respiratory symptoms and constitutional symptoms randomized patients to placebo, oseltamivir 75 mg b.i.d. or oseltamivir 150 mg b.i.d. (9, 10). While the primary endpoint was clinical resolution of symptoms, in both studies oseltamivir treatment was suggestive of decreasing nasal viral titers compared to placebo (although in neither study did this reach statistical significance). In one study this was suggested to be dosedependent (10).

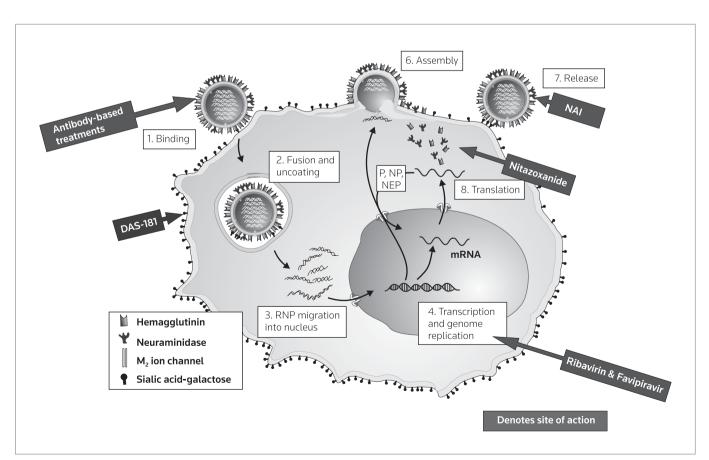


Figure 1. The influenza A virus replication cycle. Sites of novel antiviral targets are noted by the dark boxes.

Gorvorkova et al. evaluated oseltamivir and other NAIs of avian influenza viruses in a mouse model, and demonstrated a dose-related reduction in virus titers in the lungs for H5N1 and other avian influenza viruses (11). The maximum suppression was seen with the highest dose tested of 10 mg/kg/day (only tested in avian H9N2 influenza). Other murine studies indicate that higher oseltamivir doses (10 mg/kg/day instead of 1 mg/kg/day) and more prolonged administration (8 days instead of 5 days) are necessary to achieve antiviral and survival effects against a 2004 H5N1 avian influenza virus (12).

In early development studies in humans, oseltamivir dosage regimens of up to 500 mg twice daily were shown to be well tolerated (13). In an effort to determine if a higher dose of oseltamivir conveyed clinical benefit, a multicenter study was initiated in Asia and the U.S. in June 2007 to compare standard- vs. high-dose regimens (defined as 150 mg twice daily for adults with normal renal function) of oseltamivir for 5 days for severe seasonal and avian influenza. This study has recently completed enrollment, although results are not yet available (ClinicalTrials.gov Identifier NCT00298233). A separate study comparing 75 mg vs. 225 mg twice daily for 10 days in subjects with influenza in the intensive care unit was initiated in November 2009 (ClinicalTrials.gov Identifier NCT01010087). Pending the results of these studies, definitive recommendations on higher oseltamivir doses for the treatment of H1N1 influenza or other novel strains cannot be supported by clinical data.

#### Combination therapy

The paradigm of combination antivirals to more effectively control viral replication was explored in influenza long before it became an established tenant in the treatment of HIV (14). In vitro studies of dual antivirals have evaluated amantadine/oseltamivir (15-17), amantadine/ribavirin (16, 18), rimantadine/NAIs (zanamivir, oseltamivir or peramivir) (19), ribavirin/adamantanes (amantadine or rimantadine) (16-18), ribavirin/NAIs (oseltamivir or peramivir) (16, 17, 20), dual NAIs (17) and combinations involving novel agents such as favipiravir/oseltamivir (21). Dual antiviral combinations have been evaluated in animal studies, including amantadine/oseltamivir (16, 22), amantadine/ribavirin (16), rimantadine/oseltamivir (23, 24) and ribavirin/NAIs (oseltamivir and peramivir) (15, 16, 25, 26). In vivo, most dual combinations were synergistic (although some were simply additive). There is no synergy for the oseltamivir/amantadine combination for amantadine-resistant influenza virus (16). Dual NAIs have shown mixed results, with synergy shown at certain doses, while other studies demonstrated potential antagonism (17). Combination of amantadine and oseltamivir reduced the emergence of both oseltamivir- and amantadine-resistant viruses in vivo (27).

Triple antiviral combinations have been shown to increase the antiviral activity (reduction in the  $\rm EC_{50}$ ) of each drug compared to its activity in double combinations or as single agents (28). Triple antiviral combinations for viruses resistant to one component (such as amantadine resistance in 2009 H1N1) were shown to have a dose effect of that component, suggesting that incorporation of the resistant compound still contributes to antiviral efficacy (17).

Oral rimantadine plus either nebulized zanamivir or saline placebo was evaluated in the treatment of adults hospitalized with influenza. Subjects randomized to rimantadine/zanamivir were more likely

to have either no cough or only a mild cough by the third day of treatment, although there was no difference in viral shedding, duration of hospitalization, oxygen requirements or other clinical outcomes (29).

A phase II trial (ClinicalTrials.gov Identifier NCT00799760) is under way in France to compare the efficacy and safety of combination oseltamivir/zanamivir vs. monotherapy with each agent and will help determine if there is added benefit from combination NAIs. Another study comparing oseltamivir/zanamivir and oseltamivir/amantadine vs. oseltamivir monotherapy should help determine if there is clinical benefit to the synergy seen in vivo (ClinicalTrials.gov Identifier NCT00830323).

Two studies are evaluating triple antiviral therapy. The first is an international multicenter study comparing the antiviral effects of a triple-drug regimen of ADS-890 (a fixed-dose combination of amantadine and ribavirin) and oseltamivir vs. oseltamivir alone (ClinicalTrials.gov Identifier NCT00979251). A second study is evaluating amantadine/ribavirin/oseltamivir compared to NAI (oseltamivir or zanamivir) monotherapy in immunocompromised patients (ClinicalTrials.gov Identifier NCT00867139). Additional clinical studies of combination therapy are needed. Until results of these studies are available, definitive recommendations on combinations of antivirals cannot be supported by clinical data.

#### **NEW INDICATIONS FOR APPROVED DRUGS**

The use of currently licensed medications for the treatment of H1N1 would significantly shorten the development time of these new treatments. Agents in this category would likely have an established safety profile, and potentially would be off patent, which may make the medications inexpensive and therefore more widely available. Only nitazoxanide currently fits this category.

## Nitazoxanide

Nitazoxanide is a licensed antiprotozoal agent. The antiprotozoal activity of nitazoxanide is believed to be due to inhibition of pyruvic-ferredoxin oxidoreductase (POR), an enzyme essential for anaerobic energy metabolism (30). Nitazoxanide is currently licensed for the treatment of diarrhea caused by *Giardia lamblia* or *Cryptosporidium parvum*. Nitazoxanide and its metabolite tizoxanide caused concentration-dependent inhibition of influenza virus replication, with EC $_{\rm 50}$  values of 1 µg/mL for all strains tested (mean C $_{\rm max}$  for licensed dose of nitazoxanide is 10.6 µg/mL) (31). This class of agents was shown to act at the post-translational level by selectively blocking the maturation of the viral HA by impairing HA intracellular trafficking and insertion of this protein into the host plasma membrane. Based on these data, a phase II study was initiated in early 2010 for the treatment of uncomplicated influenza (ClinicalTrials.gov Identifier NCT01056380).

## **NEW FORMULATIONS OF LICENSED ANTIVIRALS**

The development of new formulations of currently licensed antivirals may shorten development times. There would be safety data on the primary compound, although different doses and  $C_{\rm max}$  may portend different toxicities. Additionally, clinicians may be familiar with the compound, increasing acceptability.

#### Zanamivir

It was first reported in 1974 that derivatives of 2-deoxy-2,3-dehydro-N-acetylneuraminic acid (DANA), a sialic acid analogue, were able to inhibit NA activity of influenza A and B viruses (32). Using a then novel computer-assisted drug design, DANA was optimized for enzyme active-site binding of the influenza NA, with the result being 4-guanidino-2-deoxy-2,3-dehydro-N-acetylneuraminic acid (zanamivir) (33). Subsequent clinical studies showed an oral bioavailability of zanamivir of only 2%. Although parenteral administration of zanamivir provided a  $C_{max}$  20 times higher than the inhaled formulation (1275 and 54  $\mu$ g/L, respectively) (34), the inhaled powder formulation proved sufficient for the treatment of outpatient influenza (35, 36).

A mouse model of influenza H5N1 suggested that viral shedding and mortality were decreased with increasing doses of zanamivir (37). Data obtained from subjects with avian influenza suggest that severe disease is associated with viremia (38). Only 17% of inhaled dry powder zanamivir is systemically absorbed (34). It is therefore unknown if inhaled zanamivir is sufficient for severe disease. Although initially developed as an inhaled powder, previous clinical studies have evaluated the efficacy of zanamivir in other formulations, including nebulized and i.v. formulations (29, 39). The recent H1N1 pandemic has reinvigorated interest in a parenteral NAI. A single-arm open-label trial is evaluating the safety data on i.v. zanamivir (ClinicalTrials.gov Identifier NCT01014988). Additionally, i.v. zanamivir is available from GlaxoSmithKline in a compassionate-use expanded-access program.

## **ANTIBODY-BASED THERAPIES**

Serum or plasma treatment of infectious diseases is a special case. Serum therapy was the first effective antimicrobial therapy, preceding the use of sulfonamides or penicillin. The safety of plasma containing influenza antibodies is reasonably well established, as current plasma supplies (for clinical indications) contain these antibodies from seasonal vaccinations and infections. Immune plasma may be collected faster than new small molecules can be developed for therapy, and plasma may therefore be considered as a therapeutic for emerging diseases (40).

The therapeutic efficacy of  $F(ab')_2$  fragments was demonstrated in a BALB/c mouse model given a lethal dose of H5N1 virus (41). In this model, the mice received an i.p. injection of 50, 100 or 200  $\mu$ g  $F(ab')_2$  fragments/mouse with normal horse antibody as a control 24 h after infection. Anti-H5N1  $F(ab')_2$  at 100 or 200  $\mu$ g gave 100% protection against death as compared to the antibody-negative control (200  $\mu$ g of nonimmune equine antibody), with 0% survival. Eleven-week-old mice injected i.p. with 350  $\mu$ L of mouse immune serum and subsequently challenged with 10  $MLD_{50}$  of A/Vietnam/1203/04 demonstrated 100% survival as compared to 8% in the saline controls (42).

Multiple small clinical trials conducted in the Soviet Union in the 1960s-1980s reported that convalescent plasma and IVIG were effective in the treatment of influenza (43-49). These studies were often small and not randomized, but consistent beneficial effects suggested that antibody-based therapies may alter the course of disease enough to afford clinical benefit. A meta-analysis of the use of plasma therapy in 1918 influenza compiled data from 8 studies

involving 1,703 patients administered convalescent plasma (50). The typical volume of plasma or serum administered was 125-250 mL. While these studies were not randomized or blinded, and the intervention was not standardized for volume or HA antibody titer, the overall crude case-fatality rate was 16% (54 of 336) among treated patients and 37% (452 of 1,219) among controls. Efforts to collect plasma with high-titer H1N1 antibodies are currently under way, and a treatment study is scheduled to be open for enrollment by mid-2010 (ClinicalTrials.gov Identifiers NCT00984451 and NCT01052480).

Plasma-based therapies, as they are collected from human donors, are relatively expensive and improved manufacturing costs are not achieved by increasing the economies of scale. Monoclonal antibodies for influenza have the potential for improved costs through economies of scale, although monoclonal antibodies are still in preclinical development and have not yet started clinical trials.

## **NEW ANTIVIRAL AGENTS**

## Peramivir

Peramivir (also known as BCX-1812 or RWJ-270201) is a highly selective inhibitor of influenza A and B virus NA. Unlike oseltamivir or zanamivir, peramivir has a cyclopentane structure and was designed for improved binding to the active site on the NA enzyme (51). While preclinical models predicted good bioavailability and once-daily dosing (52, 53), clinical studies revealed an oral bioavailability of roughly 3% (54). Large doses were able to overcome this poor bioavailability, with phase II studies showing significantly reduced viral shedding at doses of 400 mg daily (54).

Given the need for a parenteral NAI agent, clinical studies of i.v. and i.m. peramivir began in 2006. Four i.v. and two i.m. studies have been completed. While none have been published to date, they are summarized in the FDA Fact Sheet for Health Care Professionals. In uncomplicated influenza, a single i.v. dose of peramivir was better than placebo for time to alleviation of symptoms (although this was not true for a single i.m. dose) (55). In comparison to oseltamivir, peramivir did not demonstrate superiority for the primary endpoint of alleviation of symptoms. Although peramivir is still being evaluated in licensing studies, due to the pandemic with H1N1, the U.S. FDA made i.v. peramivir available through a mechanism called an Emergency Use Authorization (EUA) (56). This is the first EUA that has been issued for an unapproved drug. As this is an unapproved drug, the use of peramivir is restricted to confirmed 2009 H1N1 infections, especially in subjects who are at high risk or may not tolerate standard antivirals. The development of oseltamivir resistance due to H275Y also confers clinically significant peramivir resistance (7).

# Laninamivir and CS-8958

Like other NAIs, laninamivir (previously known as R-125489) is a sialic acid analogue that has activity against a variety of influenza A and B viruses. Laninamivir has a very similar structure to zanamivir, with a methyl substitution for hydrogen in one side-chain. The  $IC_{50}$  for laninamivir is generally 50-100% higher than that for zanamivir. However, when laninamivir is acylated with a carbon chain at position 3 (forming the prodrug CS-8958), the half-life of the active drug (laninamivir) significantly increases. In a BALB/c mouse model,

treatment with zanamivir, laninamivir or CS-8958 4 h before infection with influenza A/PR/8/34 showed 0%, 20% and 100% survival, respectively (57).

CS-8958 is currently being developed as an inhaled formulation. In a dose-escalating phase I study, a single inhaled dose of CS-8958 had a serum half-life of 1.7-29 h but gave a half-life of up to 80 h for the active compound laninamivir (at 80 mg) (58). Mouse studies suggest that CS-8958 is rapidly metabolized to laninamivir in the respiratory tract, with a lung half-life of 0.8 h (59). Laninamivir was present in the lung for several days, likely being gradually absorbed, accounting for its long half-life (40 h in mice in this study). Comparatively, laninamivir given i.v. to mice has a half-life of 0.5 h.

Daiichi Sankyo reported results from a phase III study in August 2009, although to date this study has not been published (60). There were approximately 1,000 adult patients with confirmed influenza A or B who were randomized to receive either 20 or 40 mg of laninamivir as a single inhaled dose or 75 mg of oseltamivir twice daily for 5 days. The primary endpoint of the trial was time to symptom resolution, and both doses of laninamivir were as effective as oseltamivir. A separate phase II/III study was performed in 180 children, again randomized to 20 or 40 mg of laninamivir or oseltamivir 2 mg/kg twice daily for 5 days. It was reported that there was a trend towards a faster time to alleviation of influenza symptoms in the laninamivir groups, but to date the data have not been published.

## **DAS-181**

Cell-surface sialic acids (predominantly N-acetylneuraminic acid) are the host cell receptors for influenza A and B viruses. One potential mechanism to minimize the infectivity of influenza viruses is to remove this target from cell surfaces. DAS-181 is a bacterial sialidase cloned from Actinomyces viscosus coupled with the heparin-binding sequence derived from the human protein amphiregulin to serve as a respiratory epithelium-anchoring domain (61). DAS-181 has demonstrated activity against 2009 H1N1 and H5N1 isolates, and retained activity against oseltamivir- and zanamivir-resistant viruses (62-64). In mouse models with a control group mortality of 57%, DAS-181 given 48 h after infection was 100% effective at preventing death (61). It was slightly less effective when given at 72 h after infection. Additionally, in mouse models of coinfection with influenza and Streptococcus pneumoniae, treatment with DAS-181 was associated with lower bacterial colony counts (65). Whether this represents unique properties of a sialidase or simply that lower bacterial counts are associated with lower viral titers is not known. Phase I studies have been completed although not yet reported, and phase II studies are currently under way (ClinicalTrials.gov Identifier NCT01037205).

## **Favipiravir**

Favipiravir (previously known as T-705) is a pyrazine analogue (6-fluoro-3-hydroxy-2-pyrazinecarboxamide) that inhibits influenza virus RNA polymerase activity in a concentration-dependent, GTP-competitive manner (66, 67). In a mouse model, oral administration of favipiravir prevented death to a degree comparable with oseltamivir (68). Additionally, favipiravir has shown in vivo efficacy against a range of RNA viruses, including poliovirus, rhinovirus, respiratory syncytial virus, arenaviral hemorrhagic fever, Western equine

encephalitis virus and West Nile virus (67, 69-71). A phase I clinical trial was initiated in healthy adult volunteers in January 2007 in Japan. A phase II study in infected patients was initiated in October 2009 (clinicaltrials.jp Identifier JapicCTI-090934), and a phase II study in an older population with uncomplicated influenza is set to begin in early 2010 (ClinicalTrials.gov Identifier NCT01068912).

## Poly-ICLC

Peri-exposure, nonspecific activation of innate immunity through the use of therapeutic viral mimetics or pathogen-associated molecular patterns (PAMPs), such as poly-ICLC, may provide immediate protection against a broad variety of pathogens, including influenza, while at the same time activating key elements of adaptive immunity and bridging the gap between early nonspecific and longer-term specific host defense systems. Poly-ICLC is a clinically active dsRNA consisting of two homopolymers of poly I and poly C, which is then stabilized with poly-L-lysine. Stabilization increases resistance to endogenous ribonuclease hydrolysis severalfold. Poly-ICLC may act by the induction of dsRNA-dependent 2'-5'-oligoadenylate synthase and RNA helicase p68 enzyme systems, or through Toll-like receptor TLR3 (72, 73). Early downstream effects include induction of interferon beta (IFN- $\beta$ ), interleukins IL-6 and IL-12 $\alpha$ , and TNF- $\alpha$  (74).

In a mouse model, two doses of poly-ICLC (1 mg/kg per dose) administered intranasally within 12 days prior to infection with 10 LD $_{50}$  of influenza A/PR/8 virus fully (100%) protected the mice against death (75). Poly-ICLC did not prevent infection, but virus titers by hemagglutination and plaque assays showed more than a 2  $\log_{10}$  decrease in lung homogenates of pretreated mice compared with those in the lungs of the non-pretreated group. This efficacy was demonstrated up to 14 days prior to infection.

Additionally, animal studies have demonstrated efficacy against a variety of viral pathogens, including SARS, smallpox and viral agents (76, 77). As the mechanism of action is not dependent on the virus (or even knowing the causative virus in an outbreak), topical poly-ICLC may be an effective first-response medication. This agent can be combined with specific vaccination, antiviral segregation or other intervention programs. Due to this potential, intranasal poly-ICLC has entered clinical studies. A phase I dose-finding and safety study in healthy volunteers has recently completed enrollment (ClinicalTrials.gov Identifier NCT00646152).

## **CONCLUSIONS**

In the last 5-8 years, the limited armamentarium of effective antiinfluenza agents has become evident, with multiple new strategies in development attempting to improve the treatment of influenza. Most influenza infections are self-limited, although current antivirals fail to mitigate all morbidity and a significant socioeconomic impact is still associated with this disease. As new drugs and strategies come to fruition, it will be critical to define the population where these are best utilized.

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#### **DISCLOSURES**

The author states no conflicts of interest.

#### REFERENCES

- World Health Organization. Pandemic (H1N1) 2009 update 99. Accessed April 13, 2010 at http://www.who.int/csr/don/2010\_05\_07/en/index.html.
- 2. Monto, A.S. *Vaccines and antiviral drugs in pandemic preparedness.* Emerging Infect Dis 2006, 12(1): 55-60.
- 3. Moscona, A. Oseltamivir resistance—Disabling our influenza defenses. N Engl J Med 2005, 353(25): 2633-6.
- 4. High levels of adamantane resistance among influenza A (H3N2) viruses and interim guidelines for use of antiviral agents—United States, 2005-06 influenza season. MMWR Morb Mortal Wkly Rep 2006, 55(2): 44-6.
- Update: Influenza activity—United States, September 28-November 29, 2008. MMWR Morb Mortal Wkly Rep 2008, 57(49): 1329-32.
- 6. Update: Drug susceptibility of swine-origin influenza A (H1N1) viruses, April 2009. MMWR Morb Mortal Wkly Rep 2009, 58(16): 433-5.
- 7. Memoli, M.J., Hrabal, R.J., Hassantoufighi, A., Eichelberger, M.C., Taubenberger, J.K. *Rapid selection of oseltamivir- and peramivir-resistant pandemic H1N1 virus during therapy in 2 immunocompromised hosts.* Clin Infect Dis 2010, 50(9): 1252-5.
- 8. Mayburd, A.L. *Influenza antiviral therapeutics*. Recent Pat Antiinfect Drug Discov 2010, 5(1): 64-75.
- Nicholson, K.G., Aoki, F.Y., Osterhaus, A.D. et al. Efficacy and safety of oseltamivir in treatment of acute influenza: A randomised controlled trial. Neuraminidase Inhibitor Flu Treatment Investigator Group. Lancet 2000, 355(9218): 1845-50.
- Treanor, J.J., Hayden, F.G., Vrooman, P.S. et al. Efficacy and safety of the oral neuraminidase inhibitor oseltamivir in treating acute influenza: A randomized controlled trial. US Oral Neuraminidase Study Group. JAMA 2000, 283(8): 1016-24.
- Govorkova, E.A., Leneva, I.A., Goloubeva, O.G., Bush, K., Webster, R.G. Comparison of efficacies of RWJ-270201, zanamivir, and oseltamivir against H5N1, H9N2, and other avian influenza viruses. Antimicrob Agents Chemother 2001, 45(10): 2723-32.
- 12. Yen, H.-L., Monto, A.S., Webster, R.G., Govorkova, E.A. Virulence may determine the necessary duration and dosage of oseltamivir treatment for highly pathogenic A/Vietnam/1203/04 influenza virus in mice. J Infect Dis 2005, 192(4): 665-72.
- 13. Massarella, J.W., He, G.Z., Dorr, A., Nieforth, K., Ward, P., Brown, A. *The pharmacokinetics and tolerability of the oral neuraminidase inhibitor oseltamivir* (Ro 64-0796/GS4104) in healthy adult and elderly volunteers. J Clin Pharmacol 2000, 40(8): 836-43.
- Galegov, G.A., Pushkarskaya, N.L., Obrosova-Serova, N.P., Zhdanov, V.M. Combined action of ribovirin and rimantadine in experimental myxovirus infection. Experientia 1977, 33(7): 905-6.
- Ilyushina, N.A., Hay, A., Yilmaz, N., Boon, A.C.M., Webster, R.G., Govorkova, E.A. Oseltamivir-ribavirin combination therapy for highly pathogenic H5N1 influenza virus infection in mice. Antimicrob Agents Chemother 2008, 52(11): 3889-97.

- Smee, D.F., Hurst, B.L., Wong, M.-H., Bailey, K.W., Morrey, J.D. Effects of double combinations of amantadine, oseltamivir, and ribavirin on influenza A (H5N1) virus infections in cell culture and in mice. Antimicrob Agents Chemother 2009, 53(5): 2120-8.
- 17. Nguyen, J.T., Hoopes, J.D., Le, M.H. et al. *Triple combination of amanta-dine, ribavirin, and oseltamivir is highly active and synergistic against drug resistant influenza virus strains in vitro.* PLoS One 2010, 5(2): e9332.
- Hayden, F.G., Douglas, R.G., Simons, R. Enhancement of activity against influenza viruses by combinations of antiviral agents. Antimicrob Agents Chemother 1980, 18(4): 536-41.
- Govorkova, E.A., Fang, H.-B., Tan, M., Webster, R.G. Neuraminidase inhibitor-rimantadine combinations exert additive and synergistic antiinfluenza virus effects in MDCK cells. Antimicrob Agents Chemother 2004, 48(12): 4855-63.
- Smee, D.F., Bailey, K.W., Morrison, A.C., Sidwell, R.W. Combination treatment of influenza A virus infections in cell culture and in mice with the cyclopentane neuraminidase inhibitor RWJ-270201 and ribavirin. Chemotherapy 2002, 48(2): 88-93.
- 21. Smee, D.F., Hurst, B.L., Wong, M.H., Bailey, K.W., Tarbet, E.B., Morrey, J.D., Furuta, Y. *Effects of the combination of favipiravir (T-705) and oseltamivir on influenza A virus infections in mice.* Antimicrob Agents Chemother 2010, 54(1): 126-33.
- 22. Ilyushina, N.A., Hoffmann, E., Salomon, R., Webster, R.G., Govorkova, E.A. *Amantadine-oseltamivir combination therapy for H5N1 influenza virus infection in mice*. Antivir Ther 2007, 12(3): 363-70.
- 23. Galabov, A.S., Simeonova, L., Gegova, G. Rimantadine and oseltamivir demonstrate synergistic combination effect in an experimental infection with type A (H3N2) influenza virus in mice. Antivir Chem Chemother 2006, 17(5): 251-8.
- Leneva, I.A., Roberts, N., Govorkova, E.A., Goloubeva, O.G., Webster, R.G. The neuraminidase inhibitor GS4104 (oseltamivir phosphate) is efficacious against A/Hong Kong/156/97 (H5N1) and A/Hong Kong/1074/99 (H9N2) influenza viruses. Antiviral Res 2000, 48(2): 101-15.
- Smee, D.F., Huffman, J.H., Morrison, A.C., Barnard, D.L., Sidwell, R.W. Cyclopentane neuraminidase inhibitors with potent in vitro anti-influenza virus activities. Antimicrob Agents Chemother 2001, 45(3): 743-8.
- Smee, D.F., Wong, M.-H., Bailey, K.W., Sidwell, RW. Activities of oseltamivir and ribavirin used alone and in combination against infections in mice with recent isolates of influenza A (H1N1) and B viruses. Antivir Chem Chemother 2006, 17(4): 185-92.
- 27. Ilyushina, N.A., Bovin, N.V., Webster, R.G., Govorkova, E.A. Combination chemotherapy, a potential strategy for reducing the emergence of drugresistant influenza A variants. Antiviral Res 2006, 70(3): 121-31.
- Nguyen, J.T., Hoopes, J.D., Smee, D.F. et al. Triple combination of oseltamivir, amantadine, and ribavirin displays synergistic activity against multiple influenza virus strains in vitro. Antimicrob Agents Chemother 2009, 53(10): 4115-26.
- Ison, M.G., Gnann, J.W., Nagy-Agren, S. et al., NAID Collaborative Antiviral Study Group. Safety and efficacy of nebulized zanamivir in hospitalized patients with serious influenza. Antivir Ther 2003, 8(3): 183-90.
- 30. Raether, W., Hanel, H. *Nitroheterocyclic drugs with broad spectrum activity.* Parasitol Res 2003, 90(Suppl. 1): S19-39.
- 31. Rossignol, J.F., La Frazia, S., Chiappa, L., Ciucci, A., Santoro, M.G. *Thiazolides, a new class of anti-influenza molecules targeting viral hemag-glutinin at the post-translational level.* J Biol Chem 2009, 284(43): 29798-808.
- 32. Meindl, P., Bodo, G., Palese, P., Schulman, J., Tuppy, H. *Inhibition of neu-raminidase activity by derivatives of 2-deoxy-2,3-dehydro-N-acetylneu-raminic acid.* Virology 1974, 58(2): 457-63.

- 33. von Itzstein, M., Wu, W.Y., Kok, G.B. et al. *Rational design of potent siali-dase-based inhibitors of influenza virus replication*. Nature 1993, 363(6428): 418-23.
- 34. Cass, L.M., Efthymiopoulos, C., Bye, A. *Pharmacokinetics of zanamivir after intravenous, oral, inhaled or intranasal administration to healthy volunteers*. Clin Pharmacokinet 1999, 36(Suppl. 1): 1-11.
- Hayden, F.G., Osterhaus, A.D., Treanor, J.J. et al. Efficacy and safety of the neuraminidase inhibitor zanamivir in the treatment of influenzavirus infections. GG167 Influenza Study Group. N Engl J Med 1997, 337(13): 874-80.
- Randomised trial of efficacy and safety of inhaled zanamivir in treatment of influenza A and B virus infections. The MIST (Management of Influenza in the Southern Hemisphere Trialists) Study Group. Lancet 1998, 352(9144): 1877-81
- Gubareva, L.V., McCullers, J.A., Bethell, R.C., Webster, R.G. Characterization of influenza A/HongKong/156/97 (H5N1) virus in a mouse model and protective effect of zanamivir on H5N1 infection in mice. J Infect Dis 1998, 178(6): 1592-6.
- 38. de Jong, M.D., Simmons, C.P., Thanh, T.T. et al. *Fatal outcome of human influenza A (H5N1) is associated with high viral load and hypercytokinemia*. Nat Med 2006, 12(10): 1203-7.
- Fritz, R.S., Hayden, F.G., Calfee, D.P. et al. Nasal cytokine and chemokine responses in experimental influenza A virus infection: Results of a placebocontrolled trial of intravenous zanamivir treatment. J Infect Dis 1999, 180(3): 586-93.
- Luke, T.C., Casadevall, A., Watowich, S.J., Hoffman, S.L., Beigel, J.H., Burgess, T.H. Hark back: Passive immunotherapy for influenza and other serious infections. Crit Care Med 2010, 38(4, Suppl.): e66-73.
- 41. Lu, J., Guo, Z., Pan, X. et al. Passive immunotherapy for influenza A H5N1 virus infection with equine hyperimmune globulin F(ab')2 in mice. Respir Res 2006. 7: 43
- Sandbulte, M.R., Jimenez, G.S., Boon, A.C.M., Smith, L.R., Treanor, J.J., Webby, R.J. Cross-reactive neuraminidase antibodies afford partial protection against H5N1 in mice and are present in unexposed humans. PLoS Med 2007, 4(2): e59.
- 43. Skriabina, E.A. [Specific therapy of influenza.]. Vestn Akad Med Nauk SSSR 1954, 4: 34-9.
- 44. Tushinskii, M.D., Skriabina, E.A. [Specific serotherapy of influenza.]. Sov Med 1955, 19(1): 13-9.
- Berezina, A.I. [Experience in treating influenza with dry anti-influenza serum of Smorodintsev; preliminary report.]. Sov Med 1956, 20(1): 67-8.
- 46. Ritova, V.V., Zhdanov, V.M. [On the method of serotherapy of influenza.]. Klin Med (Mosk) 1962, 40: 117-22.
- 47. Bilda, I., Schreiter, G. [Hyperimmune serotherapy of viral influenza.]. Kinderarztl Prax 1963, 31: 341-8.
- 48. Stavskaia, V.V., Ignateva, N.A. [On the treatment of influenza and influenzal pneumonia.]. Klin Med (Mosk) 1963, 41: 69-75.
- 49. Selivanov, A.A., Morozenko, M.A., Kallas, E.V., Nikitin, M.I., Nikitina, L.E. [Experimental for achievement of therapeutic sera against respiratory infections from immunized donors]. Vrach Delo 1967, 1: 104-6.
- 50. Luke, T.C., Kilbane, E.M., Jackson, J.L., Hoffman, S.L. *Meta-analysis:* Convalescent blood products for Spanish influenza pneumonia: A future H5N1 treatment? Ann Intern Med 2006, 145(8): 599-609.
- 51. Babu, Y.S., Chand, P., Bantia, S. et al. *BCX-1812 (RWJ-270201): Discovery of a novel, highly potent, orally active, and selective influenza neuraminidase inhibitor through structure-based drug design.* J Med Chem 2000, 43(19): 3482-6.
- 52. Drusano, G.L., Preston, S.L., Smee, D., Bush, K., Bailey, K., Sidwell, R.W. Pharmacodynamic evaluation of RWJ-270201, a novel neuraminidase

- inhibitor, in a lethal murine model of influenza predicts efficacy for oncedaily dosing. Antimicrob Agents Chemother 2001, 45(7): 2115-8.
- 53. Sweet, C., Jakeman, K.J., Bush, K. et al. *Oral administration of cyclopentane neuraminidase inhibitors protects ferrets against influenza virus infection*. Antimicrob Agents Chemother 2002, 46(4): 996-1004.
- 54. Barroso, L., Treanor, J., Gubareva, L., Hayden, F.G. *Efficacy and tolerability of the oral neuraminidase inhibitor peramivir in experimental human influenza: Randomized, controlled trials for prophylaxis and treatment.*Antivir Ther 2005, 10(8): 901-10.
- 55. FDA. Emergency Use Authorization of Peramivir IV. Fact Sheet for Health Care Providers. November 19, 2009.
- 56. Birnkrant, D., Cox, E. The Emergency Use Authorization of peramivir for treatment of 2009 H1N1 influenza. N Engl J Med 2009, 361(23): 2204-7.
- 57. Yamashita, M., Tomozawa, T., Kakuta, M., Tokumitsu, A., Nasu, H., Kubo, S. CS-8958, a prodrug of the new neuraminidase inhibitor R-125489, shows long-acting anti-influenza virus activity. Antimicrob Agents Chemother 2009, 53(1): 186-92.
- 58. Ishizuka, H., Yoshiba, S., Okabe, H., Yoshihara, K. Clinical pharmacokinetics of laninamivir, a novel long-acting neuraminidase inhibitor, after single and multiple inhaled doses of its prodrug, CS-8958, in healthy male volunteers. J Clin Pharmacol 2010, Epub ahead of print.
- 59. Koyama, K., Takahashi, M., Oitate, M., Nakai, N., Takakusa, H., Miura, S., Okazaki, O. CS-8958, a prodrug of the novel neuraminidase inhibitor R-125489, demonstrates a favorable long-retention profile in the mouse respiratory tract. Antimicrob Agents Chemother 2009, 53(11): 4845-51.
- Positive top line results from phase III study (MARVEL) of anti-influenza virus agent CS-8958. Daiichi Sankyo Co. News Release, Aug 10, 2009. Accessed April 13, 2010 at http://www.daiichisankyo.com/news/ detail/003245.html.
- 61. Malakhov, M.P., Aschenbrenner, L.M., Smee, D.F. et al. *Sialidase fusion protein as a novel broad-spectrum inhibitor of influenza virus infection*. Antimicrob Agents Chemother 2006, 50(4): 1470-9.
- 62. Triana-Baltzer, G.B., Gubareva, L.V., Nicholls, J.M. et al. *Novel pandemic influenza A(H1N1) viruses are potently inhibited by DAS181, a sialidase fusion protein.* PLoS One 2009, 4(11): e7788.
- 63. Chan, R.W.Y., Chan, M.C.W., Wong, A.C.N. et al. *DAS181 inhibits H5N1 influenza virus infection of human lung tissues*. Antimicrob Agents Chemother 2009, 53(9): 3935-41.
- 64. Triana-Baltzer, G.B., Gubareva, L.V., Klimov, A.I. et al. *Inhibition of neu*raminidase inhibitor-resistant influenza virus by DAS181, a novel sialidase fusion protein. PLoS One 2009, 4(11): e7838.
- Hedlund, M., Aschenbrenner, L.M., Jensen, K., Larson, J.L., Fang, F. Sialidase-based anti-influenza virus therapy protects against secondary pneumococcal infection. J Infect Dis 2010, 201(7): 1007-15.
- Furuta, Y., Takahashi, K., Kuno-Maekawa, M. et al. *Mechanism of action of T-705 against influenza virus*. Antimicrob Agents Chemother 2005, 49(3): 981-6.
- 67. Furuta, Y., Takahashi, K., Fukuda, Y. et al. *In vitro and in vivo activities of anti-influenza virus compound T-705*. Antimicrob Agents Chemother 2002, 46(4): 977-81.
- 68. Takahashi, K., Furuta, Y., Fukuda, Y. et al. *In vitro and in vivo activities of T-705 and oseltamivir against influenza virus*. Antivir Chem Chemother 2003, 14(5): 235-41.
- 69. Gowen, B.B., Wong, M.-H., Jung, K.-H. et al. *In vitro and in vivo activities of T-705 against arenavirus and bunyavirus infections*. Antimicrob Agents Chemother 2007, 51(9): 3168-76.
- Morrey, J.D., Taro, B.S., Siddharthan, V., Wang, H., Smee, D.F., Christensen, A.J., Furuta, Y. Efficacy of orally administered T-705 pyrazine

- analog on lethal West Nile virus infection in rodents. Antiviral Res 2008, 80(3): 377-9.
- 71. Julander, J.G., Smee, D.F., Morrey, J.D., Furuta, Y. *Effect of T-705 treatment on Western equine encephalitis in a mouse model*. Antiviral Res 2009, 82(3): 169-71.
- 72. Jacobs, B., Langland, J. When two strands are better than one: The mediators and modulators of the cellular responses to double-stranded RNA. Virology 1996, 219(2): 339-49.
- 73. Zhu, X., Nishimura, F., Sasaki, K. et al. *Toll like receptor-3 ligand poly-ICLC promotes the efficacy of peripheral vaccinations with tumor antigenderived peptide epitopes in murine CNS tumor models.* J Translation Med 2007, 5: 10.
- 74. Huang, C.C., Duffy, K.E., San Mateo, L.R., Amegadzie, B.Y., Sarisky, R.T., Mbow, M.L. A pathway analysis of poly(l:C)-induced global gene expres-

- sion change in human peripheral blood mononuclear cells. Physiol Genomics 2006, 26(2): 125-33.
- 75. Wong, J.P., Saravolac, E.G., Sabuda, D., Levy, H.B., Kende, M. *Prophylactic and therapeutic efficacies of poly(IC.LC) against respiratory influenza A virus infection in mice*. Antimicrob Agents Chemother 1995, 39(11): 2574-6.
- Barnard, D.L., Day, C.W., Bailey, K. et al. Evaluation of immunomodulators, interferons and known in vitro SARS-coV inhibitors for inhibition of SARS-coV replication in BALB/c mice. Antivir Chem Chemother 2006, 17(5): 275-84.
- Baron, S., Salazar, A., Pestka, S., Poast, J., Clark, B. Smallpox model: Protection by IFN and poly I:CLC despite evasive mechanisms. Int Conf Antiviral Res 2003.